

Conserving Genetic Diversity in Yellowstone Bison

Draft Bison genetics research outline (not for distribution or citation)

23 April, 2009

By G. Luikart, A. Perez-Figueroa, M.K. Schwartz, R. Wallen, J. Combs, F. Gardipee, and F.W.

Allendorf (order of collaborators is uncertain)

Introduction:

There is increasing concern that bison in Yellowstone National Park could lose genetic variation more rapidly than is acceptable to maintain current and long term fitness and adaptive potential. A major concern is that a large number of bison, 5 to 35%, are culled in most years, and this could rapidly reduce genetic variation. The culling will continue in the future in order to prevent spread of brucellosis to cattle, which could happen when bison migrate out of Yellowstone Park near cattle grazing areas (Interagency Bison Management Plan 2000). Genetic studies are needed to quantify rates of loss of variation, guide culling strategies, and set population size management goals in Yellowstone Park.

Evidence suggests that maintaining 1000 to 2000 bison in each of the two breeding groups (central and northern groups) should retain 95% of heterozygosity and allelic richness in Yellowstone bison over the next 200 years (Gross and Wang 2006). However, there is uncertainty in these numbers due to the possible existence of strong polygamy (dominant males), population genetic subdivision (i.e., distinct breeding groups), and inter-annual variation in population size due to recurrent, large-scale culls to reduce the risk of brucellosis transmission to cattle. Gross and Wang (2006) did not consider effects of polygamy, subdivision or extensive size fluctuations.

Evidence of genetic subdivision among Yellowstone bison was detected by Halbert (2003) and Gardipee (2007). Preliminary microsatellite data detected minor genetic differentiation between the two Yellowstone breeding areas, Hayden Valley and Lamar Valley ($F_{ST} = 0.01$; Gardipee et al., unpublished data). This suggests there is relatively high gene flow occurring between the breeding areas in Yellowstone.

We will use individual-based computer simulations of bison populations to assess the potential effects of polygamy and brucellosis risk management culling on rates of loss of heterozygosity and allelic variation over the next 200 years. Individual-based computer simulations can be used to evaluate rates of loss of heterozygosity and to estimate effective population size in age structured populations (e.g., Harris and Allendorf 1989). To parameterize the simulation models, we will use Yellowstone bison age-specific birth and death rates where data exist, and other bison population data (Table 1) where necessary (Brodie 2008, Geremia et al. 2009, and Yellowstone National Park unpublished data).

The effective population size (N_e) is important in wildlife management because it influences the rate of loss of genetic variation, inbreeding (mating between relatives), fixation of deleterious alleles, and ability of a population to respond to selection (Leberg 2005). N_e estimators also can provide the most sensitive molecular genetic metric for detecting genetic bottlenecks and sudden increased rates of loss of genetic variation (e.g. Luikart et al. 1999).

N_e is usually much less than the population census size (N_C). N_e is reduced below the N_C by phenomena such as unequal sex ratio, variation in reproductive success among individuals, and fluctuations in N_C through time. Most estimates of N_e suggest that it is only about 10-40% of N_C (Frankham 1995). Given the polygamous mating system of bison (in which few males mate with many females), the N_e could be at the lower end of this range. For example, a few males could dominate breeding over a few years and greatly reduce the N_e , even if the N_C remains large. Little is known about the degree of polygamy in bison, but computer simulations allow testing a range of polygamy to bracket degree of polygamy (and variance in male reproductive success) likely to occur.

To quantify our ability to monitor N_e and to detect a potential reduction of N_e , we will use computer simulations of Yellowstone bison. We will quantify the bias and precision of N_e estimators that use microsatellite DNA data, and will use allele frequencies similar to those in Yellowstone bison. Genetic marker-based N_e estimators generally provide the best estimates of effective population size and rates of loss of variation (Frankham 1995, Luikart et al. 1999; Allendorf and Ryman 2002; Schwartz et al. 2007). Several computational methods have been recently developed to estimate N_e from genetic marker data. However, they have not been thoroughly evaluated in age structured populations. We will evaluate the usefulness of recent estimators of N_e that are based on linkage (gametic) disequilibrium (Waples and Do 2008, Tallmon et al. 2008). We also will attempt to evaluate the usefulness of an N_e estimator based on temporal change in allele frequencies (Waples and Yokota 2007). These two methods (linkage disequilibrium and temporal variance method) are the two most promising methods for estimating N_e and detecting reductions of N_e in natural populations

Research Questions:

We have two main research questions. One reflects our need predict if Yellowstone bison are likely to lose more than 10% of genetic variation in the next 200 years. The second reflects our need to use DNA-based estimators of N_e to detect an unacceptable reduction of effective population size (e.g., $N_e < 100$ or 200), if such a reduction occurs. Our specific questions are as follows:

1) What percentage of the current heterozygosity & allelic diversity will be retained in Yellowstone bison over the next 200 years (~20 generations) given that fluctuations in population size due to culling will continue; Culling will likely lead to removals of 5% to 30% of individuals depending in population abundance and winter weather.

a. Do age structured, polygamy, population fluctuations, and subdivision (e.g., $F_{ST} = 0.01$) cause bison populations lose heterozygosity and allelic variation more rapidly than 10% in 200 years?

b. If extreme polygamy occurs, could Yellowstone bison lose > 10% of heterozygosity or 10% of allelic variation in 200 years? For simulations, we will consider a wide range of male reproductive success, ranging from random mating (Fisher-Wright population), polygamy (10% of males father 50% of offspring), to extreme polygamy where only ~10% of adult males father all offspring (e.g., are dominant) each year for five consecutive years (low turnover of dominance).

c. If extreme population size fluctuations occur (e.g., from the combined effects of natural mortality and management removals) could Yellowstone bison lose > 10% of heterozygosity and allelic variation in 200 years? We will consider realistic population growth rates (λ) ranging from 1.05 and 1.20. We will simulate the following fluctuations caused by culling:

- from ~4,000 animals to ~2000 animals culled each time the population size reaches 4,500 individuals
 - from 4,000 to 3,000 culled every time the population size reaches 4,000 individuals
 - to cull the population to 3,000 every year the population reaches >3,000 individuals
 - to cull the population to 2,000 every year the population reaches >3,500 individuals
- d. By how much can culling of only young individuals (0-to-3 year olds) slow the loss of variation (by maximizing the generation length interval, e.g. from approximately 10 years to 12 or 14 years)?
- e. Does targeted culling of one subpopulation influence rate of loss of variation (given F_{st} of 0.01 or 0.10)?
- f. How much does the rate of loss of allelic diversity depend on the number of alleles per locus? Do bi-allelic loci lose variation significantly slower than loci with 5, or loci with 10 alleles per locus?
- g. If the population unexpectedly declined to only 1,000 animals (and stayed at 1,000), would the population lose > 10% of heterozygosity or 10% of allelic variation in 200 years (e.g. if the decline occurred after generation 1 (at year 10)?

2) How can genetic monitoring be applied to provide for early detection of a severe reduction in N_e ?

- a. Can a reduction of N_e to approximately $N_e = 100$ be detected after approximately 10 years or 5 years following the reduction? (Note: An N_e of 100 leads to approximately a loss of 5% of heterozygosity in 100 years, assuming a generation length of 10 years). We will mainly evaluate the power and accuracy of the LD- N_e estimator of effective size, because its power is promising (Luikart et al., unpublished data) and its performance is poorly evaluated.
- b. What is the effect of age structure on bias and precision of the LD- N_e estimator of N_e ?
- c. Time permitting, we will compare the power and accuracy of the LD- N_e method to the standard temporal variance ($F-N_e$) method (e.g., Waples 1989; Luikart et al. 1999), and the temporal variance methods developed for age-structured populations (Ryman & Jorde 2007).

Methods (NEEDS REVISION)

Both questions above will be addressed by simulating bison populations in SimuPop (Peng and Kimmel 2005) and in Pedigog (Combes et al. in press) followed by analyses of output in programs Genepop, LD-Ne and programs we are developing.

An initial description of how these programs work and what type of outputs they can provide to those making recommendations for conservation.

Formatted: Highlight

Question one: What percent of variation will be maintained

We will simulate bison populations using available data on age structure (Table 1) and variance in male reproductive success. We will compute the N_e/N_c ratio and estimate rates of loss of heterozygosity and allelic diversity while simulating a stable bison population of abundance at 2,000 and at 3,500 individuals (these simulations require annual removal to maintain population abundance at a relatively static level). We will use these baseline as the genetic effects of more random removals that occur because migration to the low elevation winter ranges is correlated with population density and winter weather affects (Geremia et al 2009). We will also calculate N_e/N_c ratio and estimates of loss in heterozygosity and allelic diversity for some more realistic scenarios of removal (we will simulate removal of only 0 to 3 year olds vs. only adults, versus random removals to test the effects on generation length, N_e , and rates of loss of variation.

Comment [W1]: I think it would be more meaningful to make the adult removal scenario simulate the removal of seropositive animals that would in effect be a non-random removal since a higher percentage of older animals test positive than would younger animals. I am sure it makes the code writing much more complicated but it would make the model much more informative for management / conservation purposes.

We will simulate male reproductive success with approximately 10% of males fathering 50% of the offspring, and with annual random turnover of dominance status. For extreme polygamy, we will have 10% of males fathering approximately 50% of the offspring but these males will remain dominant (and father 50% of offspring) for 5 consecutive years before losing their dominance status.

H_e & AD (percent alleles remaining) will be monitored for the entire population at **100 loci** (50 msats [$H \approx 0.60$] and 50 SNPs [$H \approx 0.30$]). H_e will be converted into an N_e estimate using standard equations (as in Harris and Allendorf 1989). Annual outputs of age structure (Figure 1) and heterozygosity remaining, allelic diversity remaining, and the N_e -estimates will be plotted through time (Fig. 2).

Question two: Can genetic monitoring detect reduced N_e

We will monitor H_e , A, and compute N_e using recent N_e -estimators, for the simulated populations above (in Questions 1) to evaluate potential power to detect changes in N_e using on the ground monitoring over the next 30 years. Sensitive genetic monitoring methods are badly needed for bison and other species to detect when a population is likely to lose excessive genetic variation. N_e estimators are likely to provide most sensitive indicator for detection of a genetic bottleneck of an increased rate of loss of variation (Luikart et al. 1999; Unpublished data). N_e will be estimated from heterozygosity-loss, LD- N_e , the standard temporal method (Waples 1989) and perhaps the N_e -temporal method for age structured populations (Jorde & Ryman 2007).

The 'true N_e ' will be computed from the total population sample of individuals and loci for each of 100 simulation replications for each scenario. The true N_e will be computed from the heterozygosity-loss method (inbreeding N_e), the standard temporal method (variance N_e), and the Felsenstein (1971) method accounting for age structure in populations.

To assess the effects of sample size on accuracy and precision of the LD- N_e estimator a random sample of 100, 50, and 25 individuals will be collected every year for the first 10 years, and then every 3 years (~ 0.3 bison generations). In addition, every 3 years, we will sample 25, 50, and 100 from calves and 1-year olds to evaluate the accuracy and precision of LD- N_e for estimating N_b and N_e . We could also use this cohort sampling to evaluate the temporal method for age structured pops of Jorde & Ryman (2007). This sampling will allow us to evaluate the power (and accuracy) of the different methods for detecting serious reduction of N_e .

To assess the effects of number of loci on accuracy and precision of the LD- N_e estimator, we will compare estimates from 10, 20, and 40 msats; and then 20, 40, 80 SNPs.

Project Summary:

- 1) We will produce a computer model that simulates an idealized population (i.e., no overlapping generations, no substructure, equal contributions to reproduction) and evaluates the potential effects of management removals (similar to those directed by the Interagency Bison Management Plan) on genetic diversity over 100-200 years.
- 2) We will produce an alternate computer modeling that simulates the Yellowstone bison population using the best available estimates of demography, genetic diversity, age structure, population substructure, and polygamy for Yellowstone bison breeding groups

and includes management removals similar to those experienced since 1985, to evaluate potential effects on genetic diversity over 200 years.

- 3) We will conduct a comparison of results from the idealized and realistic population models to test the effects of overlapping generations (age structure), population substructure, polygamy (unequal contribution of gender and individuals to the next generation of bison), fluctuating population abundance and management removal strategies on rates of loss of genetic diversity (heterozygosity and allelic diversity).
- 4) An objective assessment based on modeling and other information of the abundance per primary breeding herd needed to preserve 90 and 95% of current level of genetic diversity values. This assessment will assume continued fluctuations in breeding herd and population abundance.
- 5) Recommendations regarding genetic monitoring (e.g., Ne estimation & statistical methods, samples, timing, locations) to ensure Yellowstone would have the statistical power to detect a genetically significant change in diversity over time.

The questions identified above would be the focal subject of one or two separate publications (e.g., one pub for each of the two main questions above).

Table 1. Age structure of the Yellowstone bison population from summer ground classification counts (unpublished data).

Year	Range	Adult Bull	2/3 Bulls	Yearling Bulls	Adult Cow	2/3 Cows	Yearling Cows	Calves	Unknown	Total
2004	Central	23	6.1	6	34.5	5.3	5.1	17.7	2.3	100
2005	Central	26.6	5.5	6.5	35	4.4	5.6	15.9	0.5	100
2006	Central	20.9	7.2	8.2	34.9	5.2	6.1	16	1.5	100
2007	Central	25.6	5.8	5.7	35.7	4.2	5.6	16.9	0.5	100
2008	Central	35.8	4.0	3.7	36.9	3.3	5.0	11.1	0.2	100.0
Mean	Central	26.4	5.7	6.0	35.4	4.5	5.5	15.5	1.0	100.0
2004	Northern	23.1	6.7	3.7	37.5	5	4.4	16.5	3.5	100.4
2005	Northern	24.2	7.5	5.9	36.4	4.4	4.7	13.3	3.7	100.1
2006	Northern	18.8	7.3	5.6	37	5.2	6.3	19.1	0.7	100
2007	Northern	15.8	5.9	7.1	38.5	4.8	5.5	21.7	0.6	99.9
2008	Northern	15.4	7.2	8.3	35.8	5.4	5.8	22.1	0.0	100.0
Mean	Northern	19.5	6.9	6.1	37.0	5.0	5.3	18.5	1.7	100.1

=

Literature cited (unfinished):

Allendorf, F.W., and N. Ryman. 2002. The role of genetics in population viability analysis. In: Population Viability Analysis. S. R. Beissinger and D. R. McCullough, editors. University of Chicago Press, Chicago, Illinois. pp. 50-85.

Brodie, J.F. A review of American bison (*Bos bison*) demography and population dynamics. WCS Working Papers ISSN 1530-4426. Online posting ISSN 1534-7389

Felsenstein, J., 1971. Inbreeding and variance effective numbers in populations with overlapping generations. *Genetics* 68:581-597.

Frankham R. 1995. Effective population size/adult population size ratios in wildlife: a review. *Genetical Research Cambridge*, 66, 95-106.

- Harris, R.B. and F.W. Allendorf 1989. Genetically Effective Population Size of Large Mammals: An Assessment of Estimators. *Conservation Biology*, 3: 181-191.
- Jorde P.E. and N. Ryman. 2007. Unbiased Estimator for Genetic Drift and Effective Population Size. *Genetics*, 177: 927-935.
- Luikart, G., W. Sherwin, B. Steele, and F.W. Allendorf. 1998. Usefulness of molecular markers for detecting population bottlenecks via monitoring genetic change. *Molecular Ecology*, 7:963-974.
- Luikart, G., J-M. Cornuet, F.W. Allendorf, and W.B. Sherwin. 1998. Distortion of allele frequency distributions provides a test for recent population bottlenecks. *J. Heredity* 89: 238-247.
- Luikart, G., J-M. Cornuet, and F.W. Allendorf. 1999. Temporal changes in allele frequencies provide estimates of population bottleneck size. *Conservation Biology*, 13:523-530.
- Peng, B. and M. Kimmel. 2005. simuPOP: a forward-time population genetics simulation environment. *Bioinformatics*, 21(18): 3686-3687.
- Schwartz, M.K., D.A. Tallmon, and G. Luikart. 1998. Review of DNA-based census and effective population size estimators. *Animal Conservation*, 1:293-299.
- Geremia, C., P. J. White, R.A. Garrott, R.W. Wallen, K.E. Aune, J. Treanor, and J. A. Fuller. Demography of Central Yellowstone Bison: Effects of Climate, Density, and Disease. Chapter 14 in *The Ecology of Large Mammals in Central Yellowstone*. Eds. R. Garrott, P. J. White and F. Watson. Elsevier. In press.
- Waples, R.S. 1989. A Generalized Approach for Estimating Effective Population Size From Temporal Changes in Allele Frequency. *Genetics* 121: 379-391.
- Waples RS and C. Do 2008. LDNE: a program for estimating effective population size from data on linkage disequilibrium. *Molecular Ecology Resources* 753-756.